# NAT2 slow acetylation, GSTM1 null genotype, and risk of bladder cancer: results from the Spanish Bladder Cancer Study and meta-analyses

Montserrat García-Closas, Núria Malats, Debra Silverman, Mustafa Dosemeci, Manolis Kogevinas, David W Hein, Adonina Tardón, Consol Serra, Alfredo Carrato, Reina García-Closas, Josep Lloreta, Gemma Castaño-Vinyals, Meredith Yeager, Robert Welch, Stephen Chanock, Nilanjan Chatterjee, Sholom Wacholder, Claudine Samanic, Montserrat Torà, Francisco Fernández, Francisco X Real, Nathaniel Rothman

#### Summary

Background Many reported associations between common genetic polymorphisms and complex diseases have not been confirmed in subsequent studies. An exception could be the association between *NAT2* slow acetylation, *GSTM1* null genotype, and bladder-cancer risk. However, current evidence is based on meta-analyses of relatively small studies (range 23–374 cases) with some evidence of publication bias and study heterogeneity. Associations between polymorphisms in other *NAT* and *GST* genes and bladder-cancer risk have been inconsistent.

Methods We investigated polymorphisms in NAT2, GSTM1, NAT1, GSTT1, GSTM3, and GSTP1 in 1150 patients with transitional-cell carcinoma of the urinary bladder and 1149 controls in Spain; all the participants were white. We also carried out meta-analyses of NAT2, GSTM1, and bladder cancer that included more than twice as many cases as in previous reports.

Findings In our study, the odds ratios for bladder cancer for individuals with deletion of one or two copies of the GSTM1 gene were  $1\cdot 2$  (95% CI  $0\cdot 8-1\cdot 7$ ) and  $1\cdot 9$  ( $1\cdot 4-2\cdot 7$ ) respectively (p for trend  $<0\cdot 0001$ ). Compared with NAT2 rapid or intermediate acetylators, NAT2 slow acetylators had an increased overall risk of bladder cancer ( $1\cdot 4$  [ $1\cdot 2-1\cdot 7$ ]) that was stronger for cigarette smokers than for never smokers (p for interaction  $0\cdot 008$ ). No significant associations were found with the other polymorphisms. Meta-analyses showed that the overall association for NAT2 was robust (p $<0\cdot 0001$ ), and case-only meta-analyses provided support for an interaction between NAT2 and smoking (p for interaction  $0\cdot 009$ ). The overall association for GSTM1 was also robust (p $<0\cdot 0001$ ) and was not modified by smoking status (p $=0\cdot 86$ ).

Interpretation The *GSTM1* null genotype increases the overall risk of bladder cancer, and the *NAT2* slow-acetylator genotype increases risk particularly among cigarette smokers. These findings provide compelling evidence for the role of common polymorphisms in the aetiology of cancer.

Relevance to practice Although the relative risks are modest, these polymorphisms could account for up to 31% of bladder cancers because of their high prevalence.

### Introduction

The inability to replicate results on many associations between common genetic polymorphisms and complex diseases has raised scepticism in this area of research.1 One of the few exceptions could be the association between the risk of bladder cancer and polymorphisms in two carcinogen-detoxification genes—NAT2 and GSTM1. However, evidence for an association relies on analyses of pooled data and meta-analyses of relatively small studies (range 23-374 patients, average about 100 per study), and concern has been raised about publication bias and heterogeneity of results.<sup>2-9</sup> Tobacco smoking is an important cause of bladder cancer, 10 and previous analyses have suggested that the relative risk from smoking is stronger for NAT2 slow acetylators than for rapid or intermediate acetylators.<sup>2,5,11</sup> This interaction is biologically plausible, since aromatic amines, which are thought to be the most important class of bladder carcinogens in tobacco smoke,12 are detoxified by NAT2.<sup>13</sup> However, epidemiological evidence for this interaction is even weaker than for the overall genotype association. Associations between bladder-cancer risk and polymorphisms in other carcinogen-detoxification genes such as NAT1 and other glutathione-S-transferases have been less frequently explored, with inconsistent results across studies.<sup>14-35</sup>

We report results on the associations of polymorphisms in *NAT* and *GST* genes with bladder-cancer risk and their interaction with cigarette smoking among participants in the Spanish Bladder Cancer Study. This study was designed to have adequate statistical power for rigorous evaluation of the proposed associations between genetic variation in *NAT2* and *GSTM1* and bladder-cancer risk. We also report meta-analyses of *NAT2*, *GSTM1*, smoking, and bladder cancer that include more than twice as many patients as in previous reports.

#### Lancet 2005; 366: 649-59

See Comment page 610

Division of Cancer **Epidemiology and Genetics** (M García-Closas MD, D Silverman ScD. M Dosemeci PhD. N Chatterjee PhD, S Wacholder PhD, C Samanic MS, N Rothman MD) and Core Genotype Facility at the Advanced Technology Center (M Yeager PhD, R Welch MBA, S Chanock MD), National Cancer Institute, Department of Health and Human Services Bethesda MD, USA; Institut Municipal d'Investigació Mèdica, Barcelona, Spain (N Malats MD. Prof M Kogevinas MD, I Lloreta MD. G Castaño-Vinyals BSc M Torà MD, F Fernández BSc, Prof F X Real MD): Department of Pharmacology and Toxicology and James Graham Brown Cancer Center, University of Louisville School of Medicine, KY, USA (Prof D W Hein PhD): Universidad de Oviedo, Oviedo, Spain (A Tardón MD): Consorci Hospitalari Parc Taulí, Sabadell, Spain (C Serra MD): Universitat Pompeu Fabra, Barcelona, Spain (C Serra, F X Real): Hospital General de Elche, Elche, Spain (A Carrato MD); Unidad de Investigación, Hospital Universitario de Canarias,

Correspondence to:
Dr Montserrat García-Closas,
Hormonal and Reproductive
Epidemiology Branch, Division of
Cancer Epidemiology and
Genetics, National Cancer
Institute, National Institutes of
Health, 6120 Executive Blvd,
Room 7076, MSC 7234,
Rockville, MD 20852-7234, USA
montse@nih.gov

La Laguna, Spain

(R García-Closas MD); and

Hospital del Mar-IMAS,

Department of Pathology,

Barcelona, Spain (J Lloreta)

	Cases (n=1150)	Controls (n=1149)
Demography		
Mean age (SD), years	66 (10)	65 (10)
Female	146 (13%)	147 (13%)
Male	1004 (87%)	1002 (87%)
Educational attainment*		
Less than primary	525 (46%)	539 (47%)
Primary and less than high school	452 (39%)	437 (38%)
At least high school	156 (14%)	154 (13%)
Other	14 (1%)	14 (1%)
Smoking status		
Never	159 (14%)	338 (29%)
Occasional	50 (4%)	88 (8%)
Regular		
Former	474 (41%)	458 (40%)
Current	467 (41%)	265 (23%)
Type of tobacco smoked†		
Blond tobacco only	92 (10%)	114 (16%)
Black tobacco only	383 (41%)	281 (39%)
Both types of tobacco	284 (30%)	194 (27%)
Unknown tobacco type	182 (19%)	132 (18%)
Unless otherwise stated, data are number education missing for three cases and five nformation on type of tobacco missing f	controls. †Defined or	

#### Methods

#### Study population

The Spanish Bladder Cancer Study is a hospital-based case-control study based in 18 hospitals in five areas in Spain (Asturias, Barcelona metropolitan area, Vallès/Bages, Alicante, and Tenerife). Eligible "cases" were aged 21–80 years and had newly diagnosed, histologically confirmed carcinoma of the urinary bladder in 1998–2001. Diagnostic slides from each patient were reviewed by a panel of expert pathologists to confirm the diagnosis and to ensure uniformity of classification criteria, based on the 1998 system of WHO and the International Society of Urological Pathology.<sup>36</sup>

Controls were selected from patients admitted to participating hospitals with diagnoses thought to be unrelated to the exposures of interest, such as tobacco use. The distribution of reasons for hospital admission was: 37% hernias, 11% other abdominal surgery, 23% fractures, 7% other orthopaedic problems, 12% hydrocoele, 4% circulatory disorders, 2% dermatological disorders, 1% ophthalmological disorders, and 3% other diseases. Controls were individually matched to the cases for age at interview within 5-year categories, sex, ethnic origin, and region. Information on known or potential risk factors for bladder cancer for cases and controls was collected by means of computer-assisted personal interviews during the hospital admission. 84% of eligible cases and 88% of eligible controls agreed to take part in the study and were interviewed. Of the 1219 cases and 1271 controls interviewed, 1188 (97%) cases and 1173 (92%) controls provided a blood or buccal-cell sample for DNA extraction. Seven cases and 11 controls were excluded because of low amounts of DNA. To limit heterogeneity, 16 cases with neoplasias of non-transitional histology and six non-white individuals (five cases, one control) were excluded from the analyses. 15 individuals (seven cases, eight controls) with missing information on smoking status and seven (three cases, four controls) with DNA quality-control difficulties were also excluded from the analyses. Thus, the final study population available for analysis was 1150 cases and 1149 controls, all of whom were white.

Participants were classified as never smokers if they had smoked fewer than 100 cigarettes in their lifetime and ever smokers otherwise. Ever smokers were further classified as regular smokers if they had smoked at least one cigarette per day for 6 months or longer and occasional smokers otherwise. We defined current smokers as those regular smokers who had smoked within a year of the reference date; individuals who had smoked regularly but who had stopped smoking more than 1 year before the reference date were defined as former smokers. Most (81%) smokers of known tobacco type reported smoking black tobacco. In addition, the risks of bladder cancer in relation to the risk for never smokers were similarly raised among smokers of black tobacco alone, smokers of black and blond tobacco, and smokers of unknown tobacco type (data not shown). These subgroups were therefore combined as known or likely black-tobacco smokers. We obtained informed consent from potential participants in accordance with the National Cancer Institute and local institutional review boards.

#### **Procedures**

DNA for genotype assays was extracted from leucocytes with the Puregene DNA Isolation Kit (Gentra Systems, Minneapolis, MN, USA) for 1107 cases and 1032 controls included in the analyses. DNA from another 43 cases and 117 controls was extracted from mouthwash samples by a standard phenol-chloroform method. Genotype assays were done at the Core Genotyping Facility of the Division of Cancer Epidemiology and Genetics, National Cancer Institute, with the TaqMan (Applied Biosystems, Foster City, CA, USA), MGB Eclipse (Epoch Biosciences, Bothel, WA, USA), or MASSArray (Sequenom, San Diego, CA, USA) assay. A description of and methods for each specific assay can be found at the National Cancer Institute SNP500Cancer website.37 Genotype assays were done for NAT1 (Ex1-88A>T rs1057126, Ex1-81A>C rs15561, V149I rs4987076, R187Q rs4986782, R187\* rs5030839, R33\*, D251V, R64W), NAT2 (K268R rs1208, G286E rs1799931, R64O rs1801279, Y94Y rs1041983, I114T rs1801280, L161L rs1799929, R197Q rs1799930), GSTM1 deletion (SNP500Cancer ID:GSTM1-02), GSTT1 deletion (SNP500Cancer ID:GSTT1-02), GSTP1 (I105V rs947894, A114V), and GSTM3 (V224I rs7483, IVS7 -30G>T rs1537234). All genotypes studied were in Hardy-Weinberg equilibrium among the control population. Duplicate quality-control samples showed 100% agreement for all but four assays (range 98.2% to 99.6%).

Information from the NAT1 and NAT2 singlenucleotide polymorphisms analysed in this study was used to assign the most likely NAT1 and NAT2 alleles previously identified in human populations.<sup>38,39</sup> Individuals homozygous for NAT2 rapid-acetylator alleles (NAT2\*4, NAT2\*11A, NAT2\*12A, NAT2\*12B, NAT2\*12C, NAT2\*13) were classified as rapid-acetylator phenotype; individuals homozygous for slow-acetylator alleles were classified as slow-acetylator phenotype, and heterozygous individuals (one rapid and one slow NAT2 allele) were classified as intermediate-acetylator phenotype. Individuals with missing information for four rare NAT1 single-nucleotide polymorphisms (R187\*, R33\*, D251V, and R64W with more than 99% homozygous wild-type individuals) were assumed to be homozygous for NAT1\*4. On the basis of previous studies, the NAT1\*10 allele was deemed to be the "at risk" allele. GSTM1 genotypes were defined as null (-/-) if a deletion was present in both copies of the gene and present if one (+/-) or none (+/+) of the copies had a deletion. The two GSTP1 (I105V and A114V) and GSTM3 (V224I and IVS7 -30G>T) polymorphisms investigated were in strong linkage disequilibrium  $(D'=1.0, R^2=0.10 \text{ and } D'=1.0, R^2=0.68, \text{ respectively}).$ Participants were classified according to the presence of three GSTP1 variants that have been found to encode functionally differing GSTP1 proteins: GSTP1\*A (105 Ile; 114 Ala), GSTP1\*B (105 Val; 114 Ala), and GSTP1\*C (105 Val; 114 Val).40

## Statistical analysis

Odds ratios, as measure of relative risk, and 95% CI were estimated from logistic regression models, with adjustment for sex, age at interview, region, and smoking status (never, occasional, former, or current). These unconditional models provided estimates similar to those from conditional logistic regression models for individually matched pairs. Interactions between genotypes and smoking habits were also investigated by the semiparametric maximum likelihood estimator method (SPMLE)41 to allow estimation of parameters under the assumption of genotype-smoking and genotype-sex independence in the source population. This assumption is supported by strong evidence from previous studies for independence of NAT2 and GSTM1 genotypes from cigarette smoking<sup>8,11,42</sup> and sex<sup>43</sup> in the control populations. Tests for multiplicative interaction were used to assess whether the genotype odds ratios within categories of smoking habits differed significantly from each other, or whether smoking odds ratios within genotype categories differed significantly from each other. When no multiplicative interactions were present, we also tested for additive interactions, because departures from the additive model can exist in the absence of multiplicative interactions and they might have biological implications under certain biological models.44 The synergy index was used as a measure of

Genotype	Cases	Controls	Odds ratio (95% CI)	p
NAT2*				
Rapid	55	66	1.0	
Intermediate	351	427	1.0 (0.7-1.5)	0.97
Slow	728	637	1.4 (0.9-2.1)	0.10
Slow vs rapid/intermediate			1.4 (1.2-1.7)	0.0002
GSTM1†				
+/+	70	107	1.0	
+/-	352	454	1.2 (0.8-1.7)	0.38
-/-	716	571	1.9 (1.4-2.7)	0.0002
Null vs present			1.7 (1.4-2.0)	< 0.0001
NAT1				
NAT1*4/NAT1*4	585	574	1.0	
NAT1*10/NAT1*4	327	326	1.0 (0.8-1.2)	0.62
NAT1*10/NAT1*10	53	42	1.2 (0.8–1.8)	0.48
GSTT1‡				
+/+	327	340	1.0	
+/-	572	533	1.2 (1.0-1.5)	0.05
-/-	230	248	1.0 (0.8-1.3)	0.90
GSTP1 I105V				
Ile/Ile	486	488	1.0	
Ile/Val	525	531	1.0 (0.8-1.2)	0.93
Val/Val	130	119	1.2 (0.9-1.5)	0.35
GSTP1 A114V§				
Ala/Ala	966	917	1.0	
Ala/Val	113	85	1.3 (1.0-1.8)	0.07
Val/Val	4	5	0.9 (0.2-3.4)	0.85
GSTP1 I105V/A114V combination¶				
GSTP1*A/GSTP1*A	456	441	1.0	
GSTP1*A/GSTP1*B	409	402	1.0 (0.8-1.2)	0.92
GSTP1*B/GSTP1*B	95	69	1.4 (1.0-1.9)	0.09
GSTP1*C/any other variant	113	90	1.3 (0.9–1.8)	0.12
GSTM3 V224I	_	-	- \ - ,	
Val/Val	565	588	1.0	
Val/Ile	472	451	1.1 (0.9-1.3)	0.30
lle/lle	92	88	1.0 (0.7-1.4)	0.89
GSTM3 IVS7 -30G>T	-		, , ,	-
GG	439	464	1.0	
GT	529	504	1.1 (0.9-1.4)	0.19
П	160	154	1.1 (0.8–1.4)	0.64

Odds ratios from conventional logistic regression models adjusted for sex, age, region, and smoking status. Missing information on NAT2 for 16 cases vs 19 controls; on NAT1 for 123 vs 124, including individuals with rare or undeterminable alleles (62 vs 83 with other NAT1 genotypes are not shown); on GSTM1 for 11 vs 17; on GSTT1 for four vs 12; on GSTP1 I05V for nine vs 11; on GSTP1 A114V for 24 vs 25; on GSTM3 V224 for 21 vs 22; and on GSTM3 IVS7 –30G~T for 22 vs 27. \*The proportions of NAT2 slow acetylators among cases with superficial tumours (Ta) grade 1, grade 2, and grade 3, tumours involving the submucosa (T1) grades 2/3, tumours infiltrating muscle (T2) grades 2/3, and metastatic tumours (T3/T4) were 64%, 65%, 65%, 67%, 61%, and 64% (p=0-72, p=0-80, p=0-90, p=0-94, respectively, compared with Ta/grade 1 and adjusted for age, region, and smoking status). †GSTM1 +/+ and +/- could not be distinguished for one case, who contributed to the estimation of odds ratio for GSTM1 present vs null genotypes. The proportions of GSTM1 null genotype among cases with superficial tumours Ta/grade 1, Ta/grade 2, and Ta/grade 3, tumours involving the submucosa (T1) grades 2/3, tumours infiltrating muscle (T2) grades 2/3, and metastatic tumours (T3/T4) were 61%, 62%, 61%, 67%, 61%, and 66% (p=0-79, p=0-93, p=0-14, p=0-80, and p=0-35, respectively, compared with Ta/grade 1 and adjusted for age, region, and smoking status). ‡GSTT1 +/+ and +/- could not be distinguished for 17 cases and 16 controls. \$Assay done only among cases and controls with blood DNA (96% of cases and 90% of controls). ¶Classified according to Ali-Osman and colleagues\* to reflect three functionally different GSTP1 variants: GSTP1\*A (105 lle; 114 Ala), GSTP1\*B (105 Val; 114 Ala), and GSTP1\*C (105 Val; 114 Val).

 $\textit{Table 2:} \ \mathsf{Odds\ ratios\ for\ the\ associations\ of\ polymorphisms\ in\ NAT\ and\ \mathsf{GST}\ genes\ and\ bladder-cancer\ risk}$ 

additive interaction and its CI was calculated by use of previously published formulae.<sup>45</sup>

We updated previous meta-analyses on *NAT2*, *GSTM1*, and bladder cancer and used similar selection criteria for studies—ie, case-control studies in the general population.<sup>48,11</sup> Relevant studies published up to February, 2005, were identified in a MEDLINE search. For studies of *NAT2*<sup>4,11</sup> and *GSTM1*<sup>8</sup> included in previously published meta-analyses, we used data from

Smoking characteristics	Frequency				Odds ratio (95% CI) for NAT2 slow genotype association by smoking	Odds ratio (95% CI) for joint NAT2 slow genotype and smoking association		p*
	NAT2 rapid/intermediate		NAT2 sl	ow	characteristic	NAT2 rapid/intermediate	NAT2 slow	
	Cases	Controls	Cases	Controls				
Smoking status†								
Never	66	131	91	199	0.9 (0.6-1.3)	1.0	0.9 (0.6-1.3)	
Ever	340	362	637	438	1.6 (1.3-1.9)	2.9 (2.0-4.2)	4.6 (3.2-6.6)	0.008
Occasional	16	37	32	48	1.4 (0.6-2.9)	1.2 (0.6-2.4)	1.6 (0.9-2.9)	0.28
Former	161	212	310	240	1.7 (1.3-2.2)	2-4 (1-6-3-7)	4.1 (2.8-6.1)	0.006
Current	163	113	295	150	1.4 (1.1-2.0)	5-2 (3-4-8-0)	7.5 (5.0-11.3)	0.05
Type of tobacco‡								
Never	66	131	91	199	0.9 (0.6-1.3)	1.0	0.9 (0.6-1.3)	
Black	284	272	553	328	1.6 (1.3-2.0)	3.6 (2.4-5.4)	5.9 (4.0-8.7)	0.005
Blond	40	52	52	61	1.2 (0.7-2.1)	2.5 (1.4-4.3)	2.9 (1.7-4.9)	0.36
Smoking intensity§								
Never	66	131	91	199	0.9 (0.6-1.3)	1.0	0.9 (0.6-1.3)	
<10	26	55	43	61	1.7 (0.9-3.2)	0.6 (0.3-1.1)	0.9 (0.5-1.8)	0.09
10-19	67	57	106	77	1.2 (0.7-1.9)	1-3 (0-7-2-6)	1.6 (0.9-3.0)	0.31
20-29	143	108	263	133	1-4 (1-0-2-0)	1.6 (0.9-2.9)	2.3 (1.3-4.1)	0.05
30-39	31	27	88	42	1.8 (0.9-3.5)	1-4 (0-6-3-0)	2.5 (1.3-4.8)	0.06
≥40	54	73	102	71	1.7 (1.1-2.8)	1.0 (0.5-2.0)	1.8 (0.9-3.3)	0.03

\*For differences between the odds ratio for NAT2 slow-acetylation genotype within strata defined by smoking characteristics compared with never smokers. This test is equivalent to testing whether the observed joint odds ratio for NAT2 slow-acetylator genotype and smoking characteristics differs from the product of the odds ratio for NAT2 slow genotype among never smokers and the smoking characteristic among NAT2 rapid/intermediate genotype. †Odds ratios are from conventional logistic regression models adjusted for sex, age, and region. ‡The p for interaction for former vs current smokers is 0.44 and for blond vs black tobacco is 0.33. Odds ratios are from conventional logistic regression models adjusted for sex, age, region, and smoking cessation (former/current). Black is for known or likely black tobacco smokers. \$Cigarettes per day. Odds ratios are from conventional logistic regression models adjusted for age, sex, region, smoking duration (<20 years, 20-29 years, 30-39 years, 40-49 years, ≥50 years), and smoking cessation (current/former). Odds ratios for NAT2 slow acetylation for different categories of smoking intensity did not differ significantly from the highest intensity category (p for interaction for categories <10, 10-19, 20-29, 30-39 cigarettes per day compared with ≥40 cigarettes per day are 0.96, 0.30, 0.58, and 0.90 respectively for total intensity).

Table 3: NAT2 slow-acetylation genotype, smoking characteristics, and bladder-cancer risk

those papers rather than the data from the original reports, with a few exceptions: for Taylor (1998) in Marcus and colleagues' meta-analysis,<sup>4,11</sup> and for Lin (1994) and Bell (1993) in Engel and colleagues' meta-

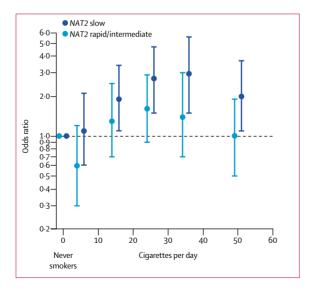


Figure 1: Association between smoking intensity (average number of cigarettes per day in categories of 10 cigarettes) and bladder-cancer risk compared with never smokers, stratified by NAT2 acetylation genotype Odds ratios are from conventional logistic regression models adjusted for age, sex, region, smoking duration (<20 years, 20-29 years, 30-39 years, 40-49 years, >50 years), and smoking cessation (current/former smokers). Error bars represent 95% Cl. p values for interaction are shown in table 3.

analysis<sup>8</sup> we used the original report to distinguish between black and white individuals; for Horai (1989) and Karakaya (1986) in Marcus and colleagues' paper,<sup>11</sup> we recalculated odds ratios and 95% CI to obtain exact estimates. For studies not included in previous meta-analyses that did not present crude odds ratios and 95% CI, we calculated them from published data.

Random-effects summary measures were calculated by weighting of each study result by a factor of withinstudy and between-study variance.46 Homogeneity of study results was assessed by the Q statistic, and publication bias was assessed by Begg's<sup>47</sup> and Egger's tests.48 A case-only design49 was used in meta-analyses to assess the presence of a multiplicative interaction between NAT2 and GSTM1 genotypes and smoking status (ever/never) because that approach meant we could include some studies without information on the cross-classification of genotype and smoking status among controls, it removed possible biases resulting from the inclusion of hospital controls with diseases related to tobacco use, and it is a powerful design to test for multiplicative interactions under the assumption of independence of NAT2 and GSTM1 from smoking status in the population. Statistical analyses were done with STATA (version 8.2, special edition).

## Role of the funding source

The study sponsors had no role in the design of the study; in the collection, analysis, or interpretation of the data; or in the writing of the report. The corresponding

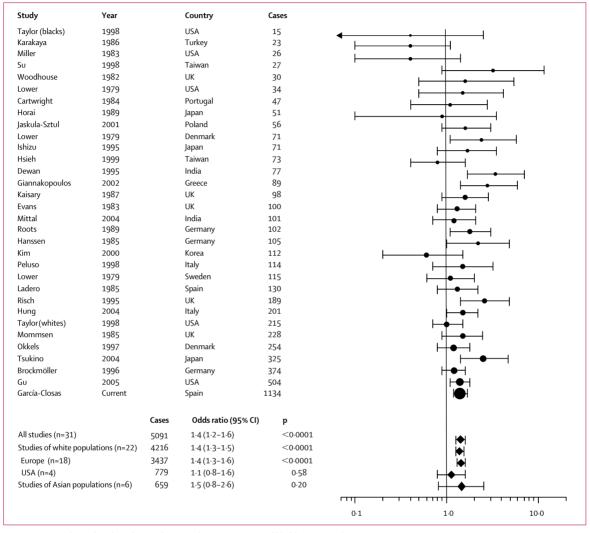


Figure 2: Meta-analysis of studies of NAT2 slow-acetylation genotype and bladder-cancer risk Numbers of cases are individuals with NAT2 information.

author had full access to all the data in the study and had final responsibility for the decision to submit the paper for publication.

## Results

The study population was white, predominantly male, and a high proportion were smokers, mostly of black tobacco (table 1). In this population, NAT2 slow-acetylator and GSTM1 null (-/-) genotypes significantly increased the risk of bladder cancer (table 2). The risk of bladder cancer was 40% higher in NAT2 slow acetylators than in NAT2 rapid or intermediate acetylators (odds ratio  $1\cdot4$  [95% CI  $1\cdot2-1\cdot7$ ]); NAT2 rapid acetylators and intermediate acetylators had similar risks of bladder cancer (table 2). The odds ratios for bladder cancer for individuals with deletion of one or two copies of the GSTM1 gene were  $1\cdot2$  ( $0\cdot8-1\cdot7$ ) and  $1\cdot9$  ( $1\cdot4-2\cdot7$ ), respectively (trend test p<0·0001). Individuals with the

null genotype had a 70% higher risk of bladder cancer than those with one or two copies of the *GSTM1* gene (table 2). The associations for *NAT2* and *GSTM1* genotypes were similar irrespective of tumour grade or stage (table 2), and there was no evidence that these associations differed by age or sex (data not shown).

The joint association for the combined *NAT2* slow-acetylator and *GSTM1* null genotype, present in 28% of the control population, compared with *NAT2* rapid/intermediate-acetylator and *GSTM1* present genotype (odds ratio  $2 \cdot 2$  [ $1 \cdot 7 - 2 \cdot 9$ ]) was consistent with a weak multiplicative interaction between these two genetic variants; however, the test for multiplicative interaction was not significant (p=0·15). None of the other genetic polymorphisms investigated was significantly associated with an increased risk of bladder cancer (table 2), and there was no evidence of multiplicative interactions between them (data not shown).

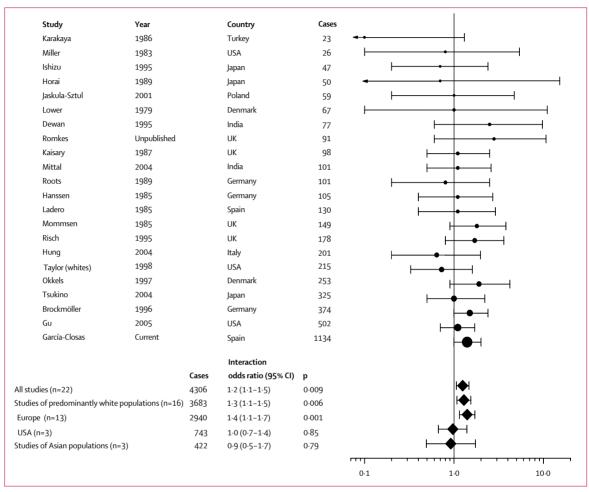


Figure 3: Case-only meta-analysis of studies of NAT2 slow-acetylation genotype, cigarette smoking, and bladder-cancer risk Numbers of cases are individuals with NAT2 and smoking information.

Conventional logistic regression analyses showed a significant multiplicative interaction between NAT2 slow acetylation and cigarette smoking status (ever/never p=0.008; table 3) with an interaction odds ratio of 1.8 $(1\cdot 2-2\cdot 8)$ . The evidence for a multiplicative interaction was somewhat weaker (interaction odds ratio 1.4 [1.0-1.9], p=0.08) when we used SPMLE logistic regression, which assumed genotype-smoking and genotype-sex independence conditional on age, in the source population. Estimates for the NAT2 slowacetylation association with bladder cancer were similar for occasional, current, and former smokers (table 3). The data suggested that the association of NAT2 slowacetylation genotype with bladder cancer was stronger for known or likely smokers of black tobacco than for smokers of blond tobacco (table 3). However, this difference was not significant (table 3). The NAT2 and smoking intensity interaction is described by showing the odds ratios for NAT2 slow acetylation genotype by smoking intensity (table 3), for the joint association of NAT2 slow genotype and smoking intensity (table 3), and for smoking intensity by *NAT2* acetylation genotype (figure 1). *NAT2* slow acetylators were at a higher risk from cigarette smoking than rapid or intermediate acetylators, for all smoking intensities (figure 1). The magnitude of the association between *NAT2* slow acetylation and bladder-cancer risk among regular smokers was similar across different smoking intensities (table 3), durations, and pack-years (data not shown). As with the interaction between *NAT2* and smoking status, SPMLE odds ratios and p values for interactions with other smoking characteristics were slightly attenuated compared with conventional analyses (data not shown).

Neither conventional nor SPMLE logistic regression showed a significant multiplicative interaction (odds ratio  $0.7 \ [0.4-1.1]$ , p=0.09, and  $0.8 \ [0.5-1.1]$ , p=0.15, respectively) for the association of *GSTM1* null and smoking status (ever/never) on bladder-cancer risk. Thus, the relative risk of bladder cancer for *GSTM1* null compared with present genotype does not vary by smoking status. No multiplicative interactions were found for other smoking characteristics such as smoking

cessation (current  $\nu$ s former smokers), smoking intensity, or duration (data not shown). Since an additive interaction can exist in the absence of a multiplicative interaction, and departures from the additive model might have biological implications under certain assumptions, we then tested for an additive interaction. Both conventional and SPMLE logistic regressions showed significant departures from the additive model (ie, additive interactions) or GSTM1 null genotype and smoking status, with synergy indices of  $1\cdot 3$  (95% CI  $1\cdot 0-1\cdot 6$ ; p=0·04) and  $1\cdot 4$  ( $1\cdot 1-1\cdot 7$ ; p=0·001), respectively.

We updated a previously published meta-analysis of 22 studies of *NAT2* and bladder cancer<sup>4</sup> to include data from our study and eight additional studies,  $^{17-19,27,28,34,50,51}$  including a total of 5091 cases and 6501 controls (figure 2). The summary relative risk for *NAT2* slow acetylators compared with rapid/intermediate acetylators was 1.4 (1.2-1.6; p<0.0001) with no

evidence for publication bias according to Begg's (p=0·94) or Egger's tests (p=0·91). There was some evidence of study heterogeneity (Q test p=0·04), which was not present when 15 studies with fewer than 100 cases each were excluded (summary odds ratio 1·4 [1·2–1·5]; Q test p=0·31). Summary estimates for white populations (56% prevalence of *NAT2* slow acetylators in controls) and Asian populations (11% prevalence of *NAT2* slow acetylators in controls) were similar (p=0·87; figure 2). The summary relative risk for studies of white populations in the USA was lower than that for studies done in Europe, which accounted for most (82%) white cases; however this difference was not significant (p=0·18; figure 2).

We also updated a case-only meta-analysis of *NAT2* and smoking interaction on bladder-cancer risk<sup>11</sup> to include results from our study and five additional studies published after the meta-analysis<sup>17,19,34,50,51</sup> (figure 3). This analysis included a total of 4306 cases and showed

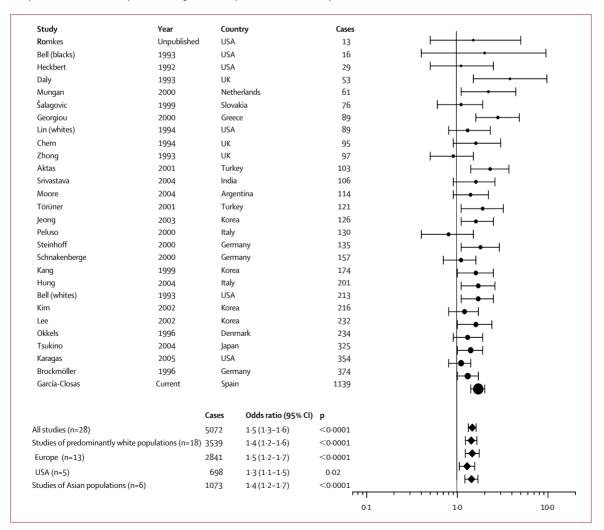


Figure 4: Meta-analysis of studies of GSTM1 null genotype and bladder-cancer risk Number of cases for studies in Engel et al $^{\rm 8}$  are based on table 1 of that paper.

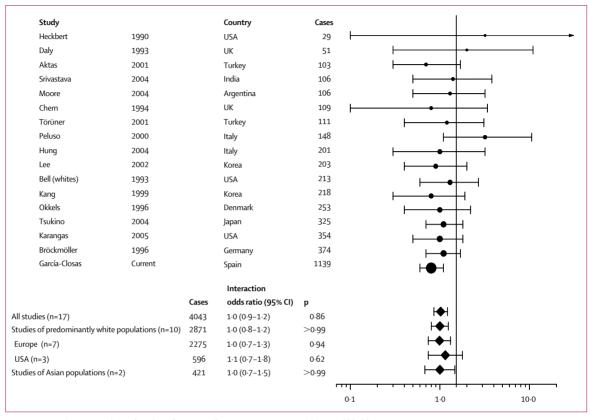


Figure 5: Case-only meta-analysis of studies of GSTM1 null genotype, cigarette smoking, and bladder cancer

Numbers of cases are individuals with GSTM1 and smoking information. Numbers of cases in studies included in Engel et al<sup>®</sup> are based on data used for the pooled analyses published in that paper.

evidence for an interaction with a summary estimate of  $1\cdot 2$   $(1\cdot 1-1\cdot 5; p=0\cdot 009)$  for all populations combined. There was no evidence of overall study heterogeneity (Q test p=0·84) or publication bias (Begg's test p=0·40, Egger's test p=0·13). The point estimate for interaction was higher in white than in Asian populations ( $1\cdot 3$  vs 0·9) and in European than in US white populations ( $1\cdot 4$  vs 1·0); however, these differences were not significant (p=0·32 and 0·09, respectively; figure 3).

A meta-analysis of 17 studies of  $GSTM1^8$  was also updated to include our study, ten additional studies, <sup>17,21,22,24,26,29,30,35,52,53</sup> and an update from a previously published study, <sup>50</sup> yielding a total of 5072 cases and 6466 controls (figure 4). The summary odds ratio for GSTM1 null versus present genotype for all populations combined was 1.5 (1.3-1.6; p<0.0001) with no evidence of study heterogeneity (Q test p=0.10) or publication bias by Begg's test (p=0.27) or Egger's test (p=0.57). Summary estimates were similar and significant in white populations (51% of GSTM1 null genotype in controls) and Asian populations (53% of GSTM1 null genotype in controls), as well as in US and European white populations (figure 4).

An updated case-only meta-analysis of studies that investigated the *GSTM1*-smoking interaction<sup>8</sup> to include

our study and seven other studies  $^{17,21,22,29,30,35,52}$  (17 studies of 4043 cases) confirmed the absence of a multiplicative interaction with a summary odds ratio of  $1 \cdot 0$  ( $0 \cdot 9 - 1 \cdot 2$ ; p=0.86; figure 5). The Q test showed no evidence of study heterogeneity (p=0.87), and Begg's test (p=0.15) and Egger's test (p=0.03) suggested the presence of publication bias. Summary estimates for the interaction were very similar for all population subgroups (figure 5).

## Discussion

This report provides compelling evidence of an increased bladder-cancer risk associated with the *GSTM1* null and *NAT2* slow-acetylation genotypes. The latter association was particularly important among cigarette smokers. Although the relative risks for polymorphisms in *NAT2* and *GSTM1* genes are modest, these polymorphisms could account for a large proportion of bladder cancers because they are very common in the population. From our data, we estimate that these polymorphisms cause 31% (95% CI 20–46) of bladder cancers in white populations. In addition, we provide strong evidence against a substantial overall association for polymorphisms in other *NAT* and *GST* genes, with the possible exception of small to moderate associations for the *NAT1\*10/\*10* and *GSTP1\*114Val/Val* genotypes.

The new meta-analysis of studies of NAT2 slow acetylation and bladder-cancer risk showed that this association is robust and similar for white and Asian populations. The lack of significance for the association in Asian populations might be explained by substantially lower statistical power to detect associations in Asian studies owing to a lower prevalence of NAT2 slow acetylators (11% for Asian vs 56% for white populations), along with a smaller number of cases available for the meta-analysis. We also found that NAT2 slow acetylators are especially susceptible to the adverse effects of cigarette smoking on bladder-cancer risk. This gene-environment interaction has strong biological plausibility, because NAT2 slow acetylators have decreased capacity to detoxify aromatic monoamines by N-acetylation,13 tobacco smoking is a primary source of exposure to aromatic amines in the general population, and aromatic amines are suspected to be the primary bladder carcinogen in tobacco smoke.<sup>12</sup> Our data suggest that NAT2 slow acetylation does not increase bladdercancer risk among never smokers, although they do not rule out a small increase in risk in this group.

Because the content of aromatic amines is higher in black than in blond tobacco,54 the effect of NAT2 slow acetylation could conceivably be stronger for smokers of black tobacco. Our data are consistent with this hypothesis, although the differences were not significant. The magnitude of the association between NAT2 slow acetylation and bladder-cancer risk was similar for different smoking intensities in our study population. Our meta-analysis of the interaction between smoking status and NAT2 slow-acetylation genotype suggested a stronger interaction with ever/ never smoking in European than in US studies (p=0.09). This difference could result from the lower content of aromatic amines in blond tobacco, which is generally smoked in the USA, than in the black tobacco commonly smoked in parts of Europe. This explanation is consistent with a study of a population in the USA that found an interaction between NAT2 slow-acetylation genotype and smoking only for heavy smokers.34

Distinction of individuals with one and two copies of the GSTM1 gene, an issue that has not been adequately addressed in previous studies of bladder cancer, suggests the presence of a gene-dosage effect with relative risks of 1.2 (0.8-1.7) and 1.9 (1.4-2.7) for individuals with one or no copies of GSTM1, respectively, compared with those with two copies (p for trend <0.0001). Meta-analyses of the association between the deletion of two copies of the GSTM1 gene (null genotype) compared with the presence of one or two copies (present genotype), as calculated from previous studies that could not distinguish between these two groups of individuals, showed that this association is robust (p<0.0001) and similar in magnitude and significant across different population subgroups.

The relative risk for *GSTM1* null genotype and bladder cancer was similar for smokers and never smokers in our study population and in meta-analysis within population subgroups. This finding suggests the presence of an additive interaction, which is supported by our data (p=0.04). This observation is compatible with equal protection by GSTM1 activity against tobacco-related and non-tobacco-related bladder cancers. This finding suggests that GSTM1 lowers the risk of bladder cancer through mechanisms that are not specific to the detoxification of polycyclic aromatic hydrocarbons in tobacco smoke. Other mechanisms of action for GSTM1 could be protection from oxidative damage through metabolism of reactive oxygen species.<sup>55</sup> Our data did not confirm previously suggested differences in risk for NAT2 slow-acetylation and GSTM1 null genotypes by tumour grade or stage at presentation.<sup>26,56–59</sup> Our findings are consistent with a potential interaction between NAT2 slow-acetylation and GSTM1 null genotypes; however, further evidence is needed to confirm this interaction.

Associations between bladder-cancer risk polymorphisms in genes encoding the NAT1 enzyme involved in the activation of aromatic amines by O-acetylation,13 and other GST enzymes that have important roles in the detoxification of polycyclic aromatic hydrocarbons and other carcinogens 60 have been less fully explored. Previous studies have provided inconsistent evidence for an association between bladder-cancer risk and NAT1\*10 alone or in combination with NAT2 slow acetylation,14-19,34 GSTT1 null alone or in combination with GSTM1 null genotype, 17,20-31,35 and GSTP1 105 Val/Val genotype. 17,21,32,33 The data from our study do not support a substantial association between GSTT1 and GSTM3 genotypes and bladder-cancer risk. We found no significant increases in bladder-cancer risk associated with polymorphisms in NAT1 or GSTP1 genes; however, our estimates did not exclude a small to moderate association for the NAT1\*10/\*10 genotype compared with the NAT1\*4/\*4 genotype or for genotypes with the GSTP1 114Val allele compared with the 114Ala/Ala genotype.

Analyses by conventional logistic regression suggested a modification of the association between risk of bladder cancer and *NAT2*, *GSTM1*, and *NAT1* genotypes by sex. However, the modifications by sex were explained by unexpected differences in the genotype distribution for male and female controls.

Our study had several strengths: high participation rates, large sample size, and high-quality information on exposure and genotype. Specifically, we made an effort to improve the precision in genotype estimation by genotyping the seven single-nucleotide polymorphisms in *NAT2* that probably account for virtually all genetic variation in white populations, <sup>61</sup> and we developed assays that successfully distinguished individuals with one or two copies of the *GSTM1* and *GSTT1* genes. We also used the SPMLE method <sup>41</sup> to increase power and reduce bias in the estimation of interactions, because of the strong

evidence from previous studies for independence of *NAT2* and *GSTM1* genotypes from cigarette smoking status<sup>8,11,42</sup> and sex<sup>43</sup> in the general population. To limit selection bias, we carefully selected controls from patients admitted for various diagnoses that were thought to be unrelated to exposures of interest, including tobacco use. Genotype frequencies among the control population were similar to those previously reported. We found no significant overall differences in genotype frequencies across control diagnoses that could have biased our results.

Although this study is the largest to date on the role of genetic polymorphisms and bladder-cancer risk and had adequate statistical power to detect modest genotype associations, the power to detect interactions was limited. Meta-analyses including previous studies improved our ability to make inferences on interactions, when there was an adequate number of previous studies with homogeneous results. A consortium of bladder-cancer studies has been formed to facilitate the pooling of comparable data on environmental and genetic risk factors across studies that will help overcome the limited power of individual studies to investigate complex interrelations.

#### Contributors

M García-Closas, N Malats, D Silverman, M Dosemeci, M Kogevinas, F X Real, and N Rothman participated in the study design, enrolment of patients, and gene selection. G Castaño-Vinyals, M Torà, F Fernández, C Samanic, A Tardón, C Serra, A Carrato, and R García-Closas participated in the study design and enrolment of patients. D W Hein, M Yeager, R Welch, and S Chanock participated in gene selection and genotyping. J Lloreta participated in the pathology review. N Chatterjee and S Wacholder participated in the statistical analyses. M García-Closas did the statistical analyses and drafted the paper with input from all investigators.

# Participating study centres in Spain

Institut Municipal d'Investigació Mèdica, Universitat Pompeu Fabra, Barcelona—Coordinating Center (M Kogevinas, N Malats, F X Real, M Sala, G Castaño, M Torà, D Puente, C Villanueva, C Murta, J Fortuny, E López, S Hernández, R Jaramillo); Hospital del Mar, Universitat Autònoma de Barcelona, Barcelona (J Lloreta, S Serrano, L Ferrer, A Gelabert, J Carles, O Bielsa, K Villadiego); Hospital Germans Tries i Pujol, Badalona, Barcelona (L Cecchini, J M Saladié, L Ibarz); Hospital de Sant Boi, Sant Boi, Barcelona (M Céspedes); Centre Hospitalari Parc Taulí, Sabadell, Barcelona (C Serra, D García, J Pujadas, R Hernando, A Cabezuelo, C Abad, A Prera, J Prat); ALTHAIA, Manresa, Barcelona (M Domènech, J Badal, J Malet); Hospital Universitario, La Laguna, Tenerife (R García-Closas, J Rodríguez de Vera, A I Martín); Hospital La Candelaria, Santa Cruz, Tenerife (J Taño, F Cáceres); Hospital General Universitario de Elche, Universidad Miguel Hernández, Elche, Alicante (A Carrato, F García-López, M Ull, A Teruel, E Andrada, A Bustos, A Castillejo, J L Soto); Universidad de Oviedo, Oviedo, Asturias (A Tardón); Hospital San Agustín, Avilés, Asturias (J L Guate, J M Lanzas J Velasco); Hospital Central Covadonga, Oviedo, Asturias (J M Fernández, J J Rodríguez, A Herrero); Hospital Central General, Oviedo, Asturias (R Abascal, C Manzano, T Miralles); Hospital de Cabueñes, Gijón, Asturias (M Rivas, M Arguelles); Hospital de Jove, Gijón, Asturias (M Díaz, J Sánchez, O González); Hospital de Cruz Roja, Gijón, Asturias (A Mateos, V Frade); Hospital Alvarez-Buylla, Mieres, Asturias (P Muntañola, C Pravia); Hospital Jarrio, Coaña, Asturias (A M Huescar, F Huergo); Hospital Carmen y Severo Ochoa, Cangas, Asturias (I Mosquera).

## Conflict of interest statement

We declare that we have no conflict of interest.

#### Acknowledgments

We thank Robert C Saal (Westat, Rockville, MD, USA), Leslie Carroll, and Jane Wang (both IMS, Silver Spring, MD, USA) for their support in study and data management; Maria Sala (Institut Municipal d'Investigació Mèdica, Barcelona, Spain) for her work in data collection; physicians, nurses, interviewers, and study participants for their efforts during fieldwork; Pam Marcus and Larry Engel from the National Cancer Institute for providing datasets for meta-analyses used in their previous publications; and the Genetic Susceptibility to Environmental Carcinogens Study (http://www.gsec.net/) for bringing together the collaborative network of investigators that contributed data used in Engel and colleagues' study,\* which enabled the update of the meta-analysis on GSTM1, smoking, and bladder cancer. This work was supported by National Cancer Institute Westat contract number N02-CP-11015, FIS/Spain 00/0745 and G03/174, and CA34627.

#### References

- Colhoun HM, McKeigue PM, Davey SG. Problems of reporting genetic associations with complex outcomes. *Lancet* 2003; 361: 865–72.
- Vineis P, Marinelli D, Autrup H, et al. Current smoking, occupation, N-acetyltransferase-2 and bladder cancer: a pooled analysis of genotype-based studies. *Cancer Epidemiol Biomarkers Prev* 2001; 10: 1249–52.
- 3 d'Errico A, Malats N, Vineis P, Boffetta P. Review of studies of selected metabolic polymorphisms and cancer. *IARC Sci Publ* 1999; 148: 323–93.
- 4 Marcus PM, Vineis P, Rothman N. NAT2 slow acetylation and bladder cancer risk: a meta-analysis of 22 case-control studies conducted in the general population. *Pharmacogenetics* 2000; 10: 115–22
- 5 Green J, Banks E, Berrington A, Darby S, Deo H, Newton R. N-acetyltransferase 2 and bladder cancer: an overview and consideration of the evidence for gene-environment interaction. Br J Cancer 2000; 83: 412–17.
- 6 Johns LE, Houlston RS. N-acetyl transferase-2 and bladder cancer risk: a meta-analysis. Environ Mol Mutagen 2000; 36: 221–27.
- 7 Johns LE, Houlston RS. Glutathione S-transferase mu1 (GSTM1) status and bladder cancer risk: a meta-analysis. *Mutagenesis* 2000; 15: 399\_404
- 8 Engel L, Taioli E, Pfeiffer R, et al. Pooled analysis and meta-analysis of glutathione S-transferase M1 and bladder cancer: a HuGE review. Am J Epidemiol 2002; 156: 95–109.
- 9 d'Errico A, Taioli E, Chen X, et al. Genetic metabolic polymorphisms and the risk of cancer: a review of the literature. *Biomarkers* 1996; 1: 149–73.
- 10 Silverman DT, Devesa SS, Moore LE, Rothman N. Bladder cancer. In: Schottenfeld D, Fraumeni JF Jr, eds. Cancer epidemiology and prevention, 3rd edn. New York: Oxford University Press (in press).
- Marcus PM, Hayes RB, Vineis P, et al. Cigarette smoking, N-acetyltransferase 2 acetylation status, and bladder cancer risk: a case-series meta-analysis of a gene-environment interaction. Cancer Epidemiol Biomarkers Prev 2000; 9: 461–67.
- 12 Vineis P, Pirastu R. Aromatic amines and cancer. Cancer Causes Control 1997; 8: 346–55.
- Hein DW. Molecular genetics and function of NAT1 and NAT2: role in aromatic amine metabolism and carcinogenesis. *Mutat Res* 2002; 506–507: 65–77.
- 14 Taylor JA, Umbach DM, Stephens E, et al. The role of N-acetylation polymorphisms in smoking-associated bladder cancer: evidence of a gene-gene-exposure three-way interaction. *Cancer Res* 1998; 58: 3603–10
- 15 Cascorbi I, Roots I, Brockmoller J. Association of NAT1 and NAT2 polymorphisms to urinary bladder cancer: significantly reduced risk in subjects with NAT1\*10. Cancer Res 2001; 61: 5051–56.
- 16 Okkels H, Sigsgaard T, Wolf H, Autrup H. Arylamine N-acetyltransferase 1 (NAT1) and 2 (NAT2) polymorphisms in susceptibility to bladder cancer: the influence of smoking. Cancer Epidemiol Biomarkers Prev 1997; 6: 225–31.
- 17 Hung RJ, Boffetta P, Brennan P, et al. GST, NAT, SULT1A1, CYP1B1 genetic polymorphisms, interactions with environmental exposures and bladder cancer risk in a high-risk population. *Int J Cancer* 2004; 110: 598–604.

- 18 Hsieh FI, Pu YS, Chern HD, Hsu LI, Chiou HY, Chen CJ. Genetic polymorphisms of N-acetyltransferase 1 and 2 and risk of cigarette smoking-related bladder cancer. Br J Cancer 1999; 81: 537–41.
- 19 Jaskula-Sztul R, Sokolowski W, Gajecka M, Szyfter K. Association of arylamine N-acetyltransferase (NAT1 and NAT2) genotypes with urinary bladder cancer risk. J Appl Genet 2001; 42: 223–31.
- 20 Sanyal S, Festa F, Sakano S, et al. Polymorphisms in DNA repair and metabolic genes in bladder cancer. *Carcinogenesis* 2004; 25: 729–34.
- 21 Toruner GA, Akyerli C, Ucar A, et al. Polymorphisms of glutathione S-transferase genes (GSTM1, GSTP1 and GSTT1) and bladder cancer susceptibility in the Turkish population. *Arch Toxicol* 2001; 75: 459–64
- 22 Srivastava DS, Kumar A, Mittal B, Mittal RD. Polymorphism of GSTM1 and GSTT1 genes in bladder cancer: a study from North India. Arch Toxicol 2004: 78: 430–34.
- 23 Brockmoller J, Cascorbi I, Kerb R, Roots I. Combined analysis of inherited polymorphisms in arylamine N-acetyltransferase 2, glutathione S-transferases M1 and T1, microsomal epoxide hydrolase, and cytochrome P450 enzymes as modulators of bladder cancer risk. Cancer Res 1996; 56: 3915–25.
- 24 Kim WJ, Kim H, Kim CH, et al. GSTT1-null genotype is a protective factor against bladder cancer. *Urology* 2002; 60: 913–18.
- 25 Schnakenberg E, Breuer R, Werdin R, Dreikorn K, Schloot W. Susceptibility genes: GSTM1 and GSTM3 as genetic risk factors in bladder cancer. Cytogenet Cell Genet 2000; 91: 234–38.
- 26 Jong JH, Jin KH, Young S, et al. Association between glutathione Stransferase M1 and T1 polymorphisms and increased risk for bladder cancer in Korean smokers. *Cancer Lett* 2003: 202: 193–99.
- 27 Kim WJ, Lee HL, Lee SC, Kim YT, Kim H. Polymorphisms of N-acetyltransferase 2, glutathione S-transferase mu and theta genes as risk factors of bladder cancer in relation to asthma and tuberculosis. J Urol 2000; 164: 209–13.
- 28 Giannakopoulos X, Charalabopoulos K, Baltogiannis D, et al. The role of N-acetyltransferase-2 and glutathione S-transferase on the risk and aggressiveness of bladder cancer. Anticancer Res 2002; 22: 3801–04.
- 29 Lee SJ, Cho SH, Park SK, et al. Combined effect of glutathione S-transferase M1 and T1 genotypes on bladder cancer risk. Cancer Lett 2002; 177: 173–79.
- 30 Moore LE, Wiencke JK, Bates MN, Zheng S, Rey OA, Smith AH. Investigation of genetic polymorphisms and smoking in a bladder cancer case-control study in Argentina. *Cancer Lett* 2004; 211: 199–207.
- 31 Katoh T, Inatomi H, Kim H, Yang M, Matsumoto T, Kawamoto T. Effects of glutathione S-transferase (GST) M1 and GSTT1 genotypes on urothelial cancer risk. Cancer Lett 1998; 132: 147–52.
- 32 Harries LW, Stubbins MJ, Forman D, Howard GC, Wolf CR. Identification of genetic polymorphisms at the glutathione S-transferase Pi locus and association with susceptibility to bladder, testicular and prostate cancer. Carcinogenesis 1997; 18: 641–44.
- 33 Ma QW, Lin GF, Chen JG, Shen JH. Polymorphism of glutathione Stransferase T1, M1 and P1 genes in a Shanghai population: patients with occupational or non-occupational bladder cancer. Biomed Environ Sci 2002; 15: 253–60.
- 34 Gu J, Liang D, Wang Y, Lu C, Wu X. Effects of N-acetyl transferase 1 and 2 polymorphisms on bladder cancer risk in Caucasians. Mutat Res 2005; 581: 97–104.
- 35 Karagas MR, Park S, Warren A, et al. Gender, smoking, glutathione-S-transferase variants and bladder cancer incidence: a populationbased study. Cancer Lett 2005; 219: 63–69.
- 36 Epstein JI, Amin MB, Reuter VR, Mostofi FK. The World Health Organization International Society of Urological Pathology consensus classification of urothelial (transitional cell) neoplasms of the urinary bladder. Am J Surg Pathol 1998; 22: 1435–48.
- 37 Parker BR, Yeager M, Staats B, et al. SNP500Cancer: a public resource for sequence validation and assay development for genetic variation in candidate genes. *Nucleic Acids Res* 2004; 3: (D528–32).
- 38 Hein DW, Grant DM, Sim E. Update on consensus arylamine N-acetyltransferase gene nomenclature. *Pharmacogenetics* 2000; 10: 291–92.
- 39 Arylamine N-acetyltransferase nomenclature committee. Arylamine N-Acetyltransferase (NAT) nomenclature. Available at www.louisville.edu/medschool/pharmacology/NAT.html (accessed May 23, 2005).

- 40 Ali-Osman F, Akande O, Antoun G, Mao JX, Buolamwini J. Molecular cloning, characterization, and expression in Escherichia coli of full-length cDNAs of three human glutathione S-transferase Pi gene variants: evidence for differential catalytic activity of the encoded proteins. J Biol Chem 1997; 272: 10004–12.
- 41 Chatterjee N, Carroll RJ. Semiparametric maximum likelihood estimation exploiting gene-environment independence in casecontrol studies. *Biometrika* (in press).
- 42 Smits KM, Benhamou S, Garte S, et al. Association of metabolic gene polymorphisms with tobacco consumption in healthy controls. *Int J Cancer* 2004; 110: 266–70.
- 43 Garte S, Gaspari L, Alexandrie A-K, et al. Metabolic gene polymorphism frequencies in control populations. Cancer Epidemiol Biomarkers Prev 2001; 10: 1239–48.
- 44 Thompson WD. Effect modification and the limits of biological inference from epidemiologic data. J Clin Epidemiol 1991; 44: 221–32
- 45 Hosmer DW, Lemeshow S. Confidence interval estimation of interaction. *Epidemiology* 1992; 3: 452–56.
- 46 Laird NM, Mosteller F. Some statistical methods for combining experimental results. Int J Technol Assess Health Care 1990; 6: 5–30.
- 47 Begg CB, Mazumdar M. Operating characteristics of a rank correlation test for publication bias. *Biometrics* 1994; 50: 1088–101.
- 48 Egger M, Davey SG, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ* 1997; **315**: 629–34.
- 49 Umbach DM, Weinberg CR. Designing and analysing case-control studies to exploit independence of genotype and exposure. Stat Med 1997; 16: 1731–43.
- 50 Tsukino H, Nakao H, Kuroda Y, et al. Glutathione S-transferase (GST) M1, T1 and N-acetyltransferase 2 (NAT2) polymorphisms and urothelial cancer risk with tobacco smoking. Eur J Cancer Prev 2004; 13: 509–14.
- 51 Mittal RD, Srivastava DS, Mandhani A. NAT2 gene polymorphism in bladder cancer: a study from North India. Clin Urol 2004; 30: 270, 88
- 52 Aktas D, Ozen H, Atsu N, Tekin A, Sozen S, Tuncbilek E. Glutathione S-transferase M1 gene polymorphism in bladder cancer patients. a marker for invasive bladder cancer? *Cancer Genet Cytogenet* 2001; 125: 1–4.
- 53 Steinhoff C, Franke KH, Golka K, et al. Glutathione transferase isozyme genotypes in patients with prostate and bladder carcinoma. *Arch Toxicol* 2000; 74: 521–26.
- 54 Bartsch H, Malaveille C, Friesen M, Kadlubar FF, Vineis P. Black (air-cured) and blond (flue-cured) tobacco cancer risk: IV, molecular dosimetry studies implicate aromatic amines as bladder carcinogens. Eur J Cancer 1993; 29A: 1199–207.
- 55 Hayes JD, Strange RC. Glutathione S-transferase polymorphisms and their biological consequences. *Pharmacology* 2000; 61: 154–66.
- 56 Hanssen HP, Agarwal DP, Goedde HW, et al. Association of N-acetyltransferase polymorphism and environmental factors with bladder carcinogenesis: study in a north German population. Eur Urol 1985; 11: 263–66.
- 57 Kaisary A, Smith P, Jaczq E, et al. Genetic predisposition to bladder cancer: ability to hydroxylate debrisoquine and mephenytoin as risk factors. Cancer Res 1987; 47: 5488–93.
- 58 Inatomi H, Katoh T, Kawamoto T, Matsumoto T. NAT2 gene polymorphism as a possible marker for susceptibility to bladder cancer in Japanese. *Int J Urol* 1999; 6: 446–54.
- 59 Georgiou I, Filiadis IF, Alamanos Y, Bouba I, Giannakopoulos X, Lolis D. Glutathione S-transferase null genotypes in transitional cell bladder cancer: a case-control study. Eur Urol 2000; 37: 660-64
- 60 Hayes JD, Pulford DJ. The glutathione S-transferase supergene family: regulation of GST and the contribution of the isoenzymes to cancer chemoprotection and drug resistance. Crit Rev Biochem Mol Biol 1995; 30: 445–600.
- 61 Deitz AC, Rothman N, Rebbeck TR, et al. Impact of misclassification in genotype-exposure interaction studies: example of Nacetyltransferase 2 (NAT2), smoking, and bladder cancer. Cancer Epidemiol Biomarkers Prev 2004; 13: 1543–46.